

Published on Web 04/12/2006

## Searching for Cyclin-Dependent Kinase Inhibitors Using a New Variant of the Cope Elimination

Roger J. Griffin,\*,† Andrew Henderson,† Nicola J. Curtin,‡ Aude Echalier,§ Jane A. Endicott,§ Ian R. Hardcastle, David R. Newell, Martin E. M. Noble, Lan-Zhen Wang, and Bernard T. Golding\*,†

Northern Institute for Cancer Research, School of Natural Sciences—Chemistry, Bedson Building, University of Newcastle, Newcastle upon Tyne, NEI 7RU, U.K., Northern Institute for Cancer Research, Paul O'Gorman Building, Medical School, Framlington Place, University of Newcastle, Newcastle upon Tyne, NE2 4HH, U.K., and Laboratory of Molecular Biophysics and Department of Biochemistry, University of Oxford, Oxford, OX1 3QU, U.K.

Received January 25, 2006; E-mail: b.t.golding@ncl.ac.uk

The defective functioning of cyclin-dependent kinases (CDKs) compromises normal cell cycle progression<sup>1,2</sup> and is associated with the molecular pathology of cancer.<sup>3,4</sup> During late  $G_1$ , the CDK2/ cyclin E complex sustains hyperphosphorylation of the retinoblastoma tumor suppressor protein (pRb), which controls levels of the transcription factor E2F. Once cells enter S phase, CDK2/cyclin A phosphorylates and consequently deactivates E2F. Deregulated and elevated levels of E2F transcriptional activity lead to cell death via apoptosis. This suggests that inhibition of CDK2 may have a therapeutic benefit by eliciting tumor cell apoptosis.<sup>5</sup> Numerous CDK inhibitors have been reported,<sup>6,7</sup> the most common being those selective for CDK2 and CDK4. In this paper, we describe a synthetic strategy that has been developed for use in a multiple-parallel format and is applicable for generating a variety of compound libraries. This has enabled the discovery of new, potent inhibitors of CDK2.

With the finding that O<sup>6</sup>-cyclohexylmethylguanine 1a (Figure 1) is a moderate inhibitor of CDK1 and CDK2  $[K_i = 12 \text{ and } 5]$ μM, respectively),8 a rational design approach was implemented guided by the crystal structure of 1a bound to monomeric CDK2.9 These studies suggested that introduction of arylamino groups at C-2 of the purine, especially those substituted at the 4-position, would confer enhanced potency. We found that sulfonyl [e.g., 1b;  $IC_{50}$  (CDK2) = 63 nM] and sulfonamido substituents [e.g., 1c;  $IC_{50}$ (CDK2) = 5 nM] enhanced activity. Crystallographic studies of 1c bound to fully activated CDK2/cyclin A revealed that the enhanced potency relative to 1a is primarily a result of two additional hydrogen bonds to Asp86 and stacking interactions that these bonds promote between the arylamino ring of 1c and the CDK2 backbone.9 The sulfonamido oxygen of 1c accepts a hydrogen bond from the backbone NH of Asp86, while a sulfonamido NH atom donates a hydrogen bond to the side chain carboxylate of Asp86. Examination of the X-ray structure suggested that N-substituents on the sulfonamido group would protrude toward the surface of the C-terminal domain of CDK2. Further exploration in this region, which differs among CDKs, was expected to lead to inhibitors with enhanced selectivity for CDK2, as well as improved pharmaceutical proper-

Whereas elaboration of the sulfonamide of 1c was relatively straightforward, 10 an innovative approach was required to enable the generation of libraries of sulfone-based compounds. To achieve this goal, we conceived a procedure whereby oxidation of the  $\beta$ -piperidinoethylsulfide 2 with >3 molar equiv of 3-chloroper-

**Scheme 1.** Synthesis of  $\beta$ -Aminoethylsulfone-Based Inhibitors of CDK2 **5a**-**5w**<sup>a</sup>

<sup>a</sup> e.g. **5b**,  $R^1R^2 = CH_2CH_2NMeCH_2CH_2$ ; **5e**,  $R^1 = R^2 = Et$ ; **5h**,  $R^1$ 3-hydroxypropyl,  $R^2 = H$ ; 51,  $R^1 = 2$ -acetamidoethyl,  $R^2 = H$ ; 50,  $R^1R^2 =$  $CH_2CH_2CH(CH_2CH_2OH)CH_2CH_2$ ; **5s**,  $R^1 = 2$ -dimethylaminoethyl,  $R^2 =$ Me; **5t**,  $R^1R^2 = (CH_2)_6$ ; **5w**,  $R^1 = R^2 = 2$ -hydroxyethyl.

Scheme 2. Model Reactions with 4-(2-Piperidin-1-ylethylsulfanyl)phenyl] Carbamic Acid tert-Butyl Ester 6 Leading to 9a

<sup>a</sup> e.g.  $\mathbf{9a}$ ,  $R^1 = 2$ -hydroxyethyl,  $R^2 = H$ ;  $\mathbf{9b}$ ,  $R^1R^2 = CH_2CH_2OCH_2CH_2$ ; **9c**,  $R^1$  = cyclopentyl,  $R^2$  = H.

benzoic acid would give N-oxide sulfone 3. Cope-type elimination<sup>11</sup> of 3 would afford vinylsulfone 4 that could be captured by a variety of amines leading to products 5 (Scheme 1). The planned chemistry was initially established using [4-(2-piperidin-1-ylethylsulfanyl)phenyl] carbamic acid tert-butyl ester 6 as a model substrate. Triple oxidation of 6 resulted in the corresponding tri-oxygenated species (7), which spontaneously decomposed to N-hydroxypiperidine and vinyl sulfone 8, the structure of which was confirmed by X-ray analysis.<sup>12</sup> When vinyl sulfone 8 was mixed with N-hydroxypiperidine, 7 was partially regenerated, showing that these compounds are in equilibrium (ca. 1:1 ratio of 7 and 8 according to LC-MS). However, addition of an amine to the mixture gave the corresponding adducts 9a-9e in excellent yield (77-91%) and purity (Scheme 2 and Supporting Information). These results confirmed the

School of Natural Sciences-Chemistry, University of Newcastle.

<sup>‡</sup> Medical School, University of Newcastle. § University of Oxford.

**Scheme 3.** Synthesis of  $\beta$ -Piperidinoethylsulfide  $2^a$ 

<sup>a</sup> Reagents and conditions: (a) (i) Boc<sub>2</sub>O, NEt<sub>3</sub>, dioxane/water, rt, (ii) 3 M HCl; (b) (i) SOCl2, DMF, THF, (ii) piperidine, THF, rt; (c) 13, TFA, TFE, reflux; (d) LiAlH<sub>4</sub>, THF, rt.

$$R = H (1a) \qquad R = Me^{-S} \qquad (1b) \qquad R = H_2N \qquad (1c)$$

Figure 1. Lead CDK2 inhibitors identified by a structure-based design strategy.

existence of the equilibrium described, while providing a basis for the application of similar chemistry to the purine scaffold.

We were gratified to find that the chemistry planned for 2 proceeded exactly as intended to provide sulfones 5a-5w (Scheme 1 and Supporting Information). To access the  $\beta$ -piperidinoethylsulfide 2 (Scheme 3), commercially available 4-aminophenylthioacetic acid (10) was N-protected with t-butoxycarbonyl (Boc) to give 11, which was converted into amide 12 by reaction of an intermediate acid chloride with piperidine. On heating 12 with fluoropurine 13 under the trifluoroacetic acid/2,2,2-trifluoroethanol protocol previously described, <sup>13</sup> in situ removal of the Boc group released aniline 14, which reacted with 13 to furnish the amide 15. Reduction of 15 afforded the corresponding tertiary amine 2. Application of the methodology developed with 6 to the purine 2 (Scheme 1) gave compounds 5a-5c in acceptable yields (>65%) and sufficiently pure after HPLC for biological evaluation. The methodology was then implemented in a multiple-parallel format, allowing the synthesis of compounds 5d-5w in high purity and satisfactory yields. The "one-pot" procedure developed is applicable to numerous scenarios in the context of drug development.

Inhibitors **5a**–**5w** displayed a range of activities against CDK2, the structure-activity relationship (SAR) appearing to favor the incorporation of smaller, less sterically hindered groups (e.g., 5h;  $IC_{50} = 45$  nM). Increasingly bulky groups (e.g., **5t**;  $IC_{50} = 1.78$ μM) largely abolished activity. The structure of the CDK2/cyclin A/5h complex provided a starting point from which to rationalize the observed SAR (Figure 2). There are three hydrogen bonds between the guanine N9, N3, and the NH of the C2-substituted anilino group and the backbone carbonyl of Glu81 and the amide NH and carbonyl group of Leu83, respectively.8-10 The position of the anilino group is similar to that of 1c bound to CDK2/cyclin A<sup>9</sup> and enables the 3-hydroxypropyl group of the aminoethylsulfonyl substituent to form multiple polar contacts, notably with Asp86 and Lys89. Mimicking the interactions of the 1c sulfonamide group, the backbone amide NH and side chain carboxylate of Asp86 can hydrogen bond to a sulfone oxygen and the NH of the ethylamino group, respectively. The other sulfone oxygen is then

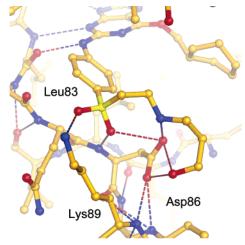


Figure 2. Structure of 5h bound to CDK2/cyclin A. Dotted lines represent polar contacts <3.4 Å.

positioned to interact with the  $\epsilon$ -NH<sub>2</sub> group of Lys89 (the first observation of such an interaction in this series) and the terminal OH group with the side chain of Asp86. All these interactions result in the extended anilino substituent adopting an ordered conformation on the CDK2 surface. However, the lower potency for 5h compared to that of 1c suggests that this network of polar contacts is insufficient to compensate for the loss of more favorable interactions between CDK2 and the purine and anilino rings of 1c. An overlay of the two inhibitors bound to CDK2/cyclin A showed that their relative binding orientations do differ.

Application of the methodology described to the development of inhibitors of other CDKs is in progress.

Acknowledgment. We thank Cancer Research UK, BBSRC (A.H.), and EU FP 6 (A.E.) for financial support.

Supporting Information Available: Details of preparative procedures/spectroscopic data for compounds 5a-5w and 9a-9d, CDK2 inhibitory data for 5a-5w; crystallization of a T160pCDK2/cyclin A/5h complex, X-ray crystallography data collection and processing; complete refs 8-10. The coordinates of the CDK2/cyclin A/5h complex have been deposited in the Protein Data Bank under ID code 2G9X. This material is available free of charge via the Internet at http://pubs.acs.org.

## References

- (1) Sherr, C. J. Science 1996, 274, 1672-1677.
- Nurse, P.; Masui, Y.; Hartwell, L. Nat. Med. 1998, 4, 1103-1106.
- (3) Fischer, P. M.; Borradori, A. G. Expert Opin. Invest. Drugs 2005, 14,
- (4) Dancey, J.; Sausville, E. A. Nat. Rev. Drug Discovery 2003, 2, 296-
- (5) Fischer, P. M. Cell Cycle 2004, 3, 742-746.
- (6) Knockaert, M.; Greengard, P.; Meijer, L. Trends Pharmacol. Sci. 2002,
- Hardcastle, I. R.; Griffin, R. J.; Golding, B. T. Annu. Rev. Pharm. Toxicol. **2002**, 42, 325

- Arris, C.; et al. J. Med. Chem. 2000, 43, 2797–2804.
  Davies, T. G.; et al. Nat. Struct. Biol. 2002, 9, 745.
  Hardcastle, I. R.; et al. J. Med. Chem. 2004, 47, 3710–3722.
- (11) Cope, A. C.; Foster, T. T.; Towle, P. H. J. Am. Chem. Soc. 1949, 71,
- (12) Henderson, A.; Clegg, W. Unpublished result at the University of Newcastle.
- Whitfield, H. J.; Griffin, R. J.; Hardcastle, I. R.; Henderson, A.; Meneyrol, J.; Mesguiche, V.; Sayle, K. L.; Golding, B. T. J. Chem. Soc., Chem. Commun. 2003, 2802-2803.

JA060595J